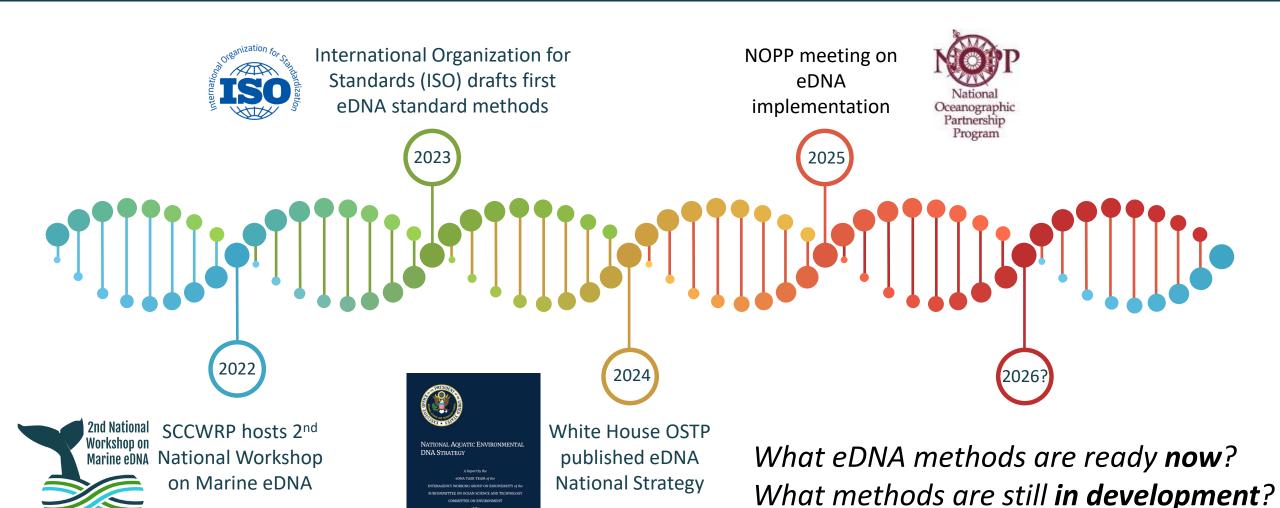
Environmental DNA (eDNA) Implementation in California

Susanna Theroux, PhD

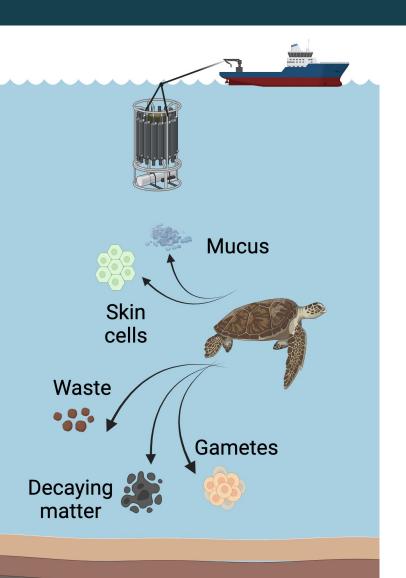




eDNA: research to implementation



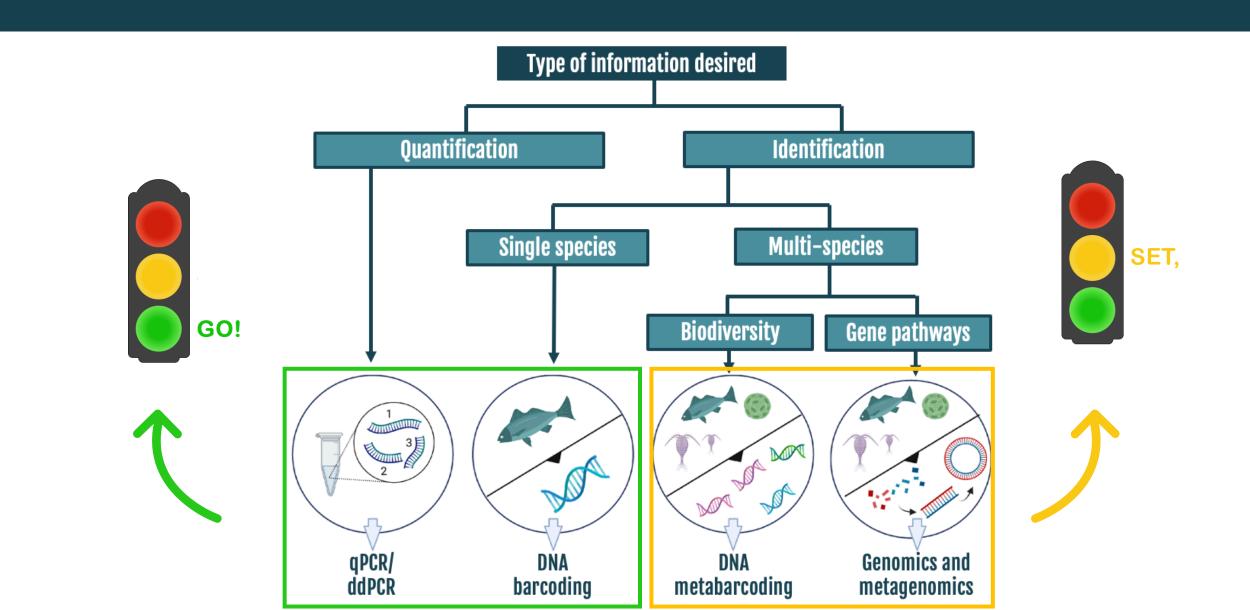
What is environmental DNA (eDNA)?





Typical eDNA workflows involve the collection of an environmental sample, filtration or precipitation to concentration the sample, DNA extraction, and analysis for either *quantification* or *identification*.

The eDNA toolkit



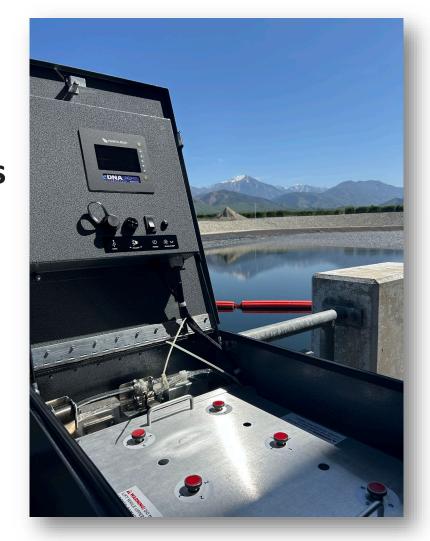
eDNA in action: Delta Smelt



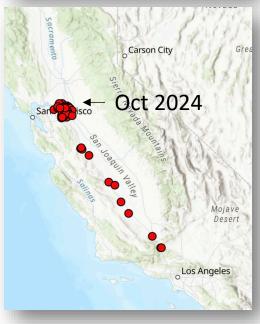
- Delta Smelt (Hypomesus transpacificus) are small, rare, short-lived, and endangered
- eDNA is non-destructive, sensitive, and enables monitoring in otherwise difficult to sample habitats
- CA DWR is using eDNA for permit compliance surveillance of Delta Smelt at pumping facility
- DWR uses qPCR and a rapid, fieldbased assay to inform management decisions

eDNA in action: Golden Mussel

- Invasive Golden Mussel (Limnoperna fortunei)
- eDNA is sensitive and able to distinguish closely related species
- Statewide eDNA monitoring network now in place with intercalibration in development
- SCCWRP is working with San Bernadino Valley MWD to test for Golden mussel in the Citrus Reservoir







eDNA in action: routine monitoring

- RB9 has been piloting ddPCR assays for invasive and endangered species monitoring
 - Water eDNA samples are collected alongside traditional bioassessment sampling
- eDNA is non-destructive, sensitive, helps expand suite of monitored taxa
- Positive detections are flagged for follow-up with additional visual and eDNA sampling





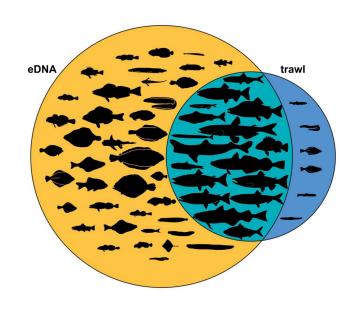






In development: community-based applications

Fish biodiversity



- Non-destructive sampling
- Ability to resolve difficult to ID taxa
- Scale across space/time

Algal Bioassessment



- Alleviates taxonomic capacity limitations
- Cost <<< traditional approaches

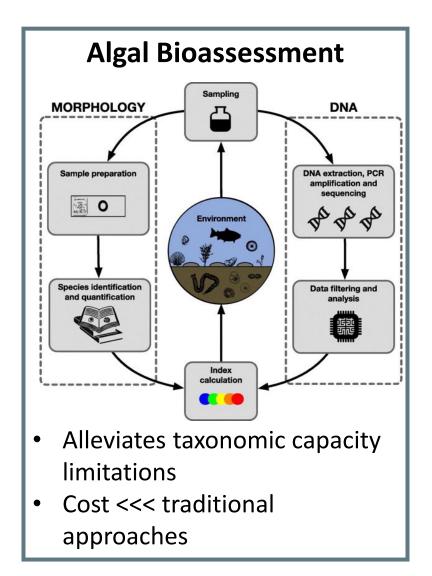
Harmful algal blooms

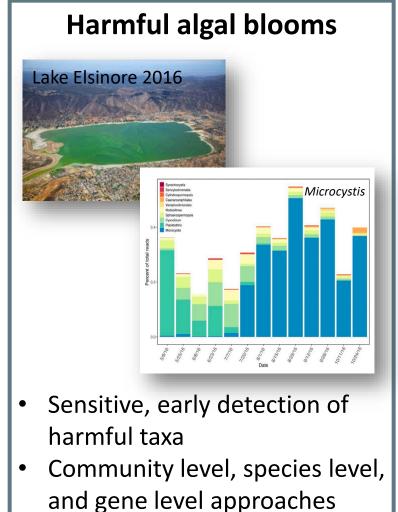


- Sensitive, early detection of harmful taxa
- Community level, species level, and gene level approaches

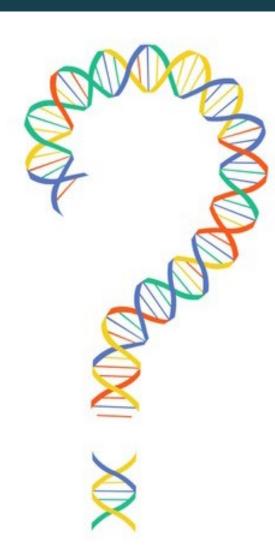
In development: community-based applications

Fish biodiversity **Monitoring Program** Non-destructive sampling Ability to resolve difficult to ID taxa Scale across space/time



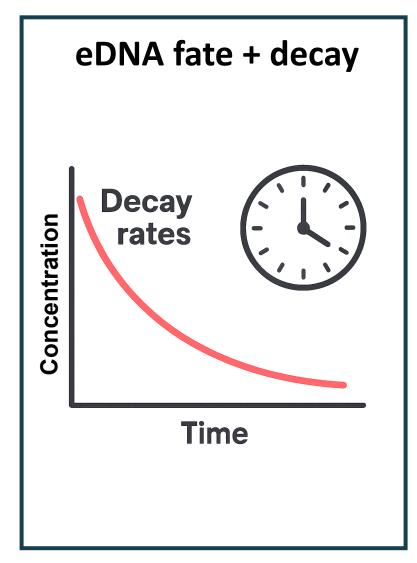


Priority eDNA research questions

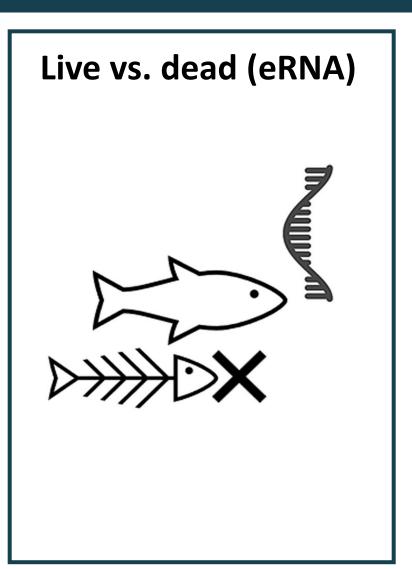


- How recently was this organism here?
- How far did the eDNA signal travel?
- Will you miss certain species because of gaps in DNA reference libraries?
- How do you know the organism is alive or dead?
- How do you know if the toxin genes are turned on or off?

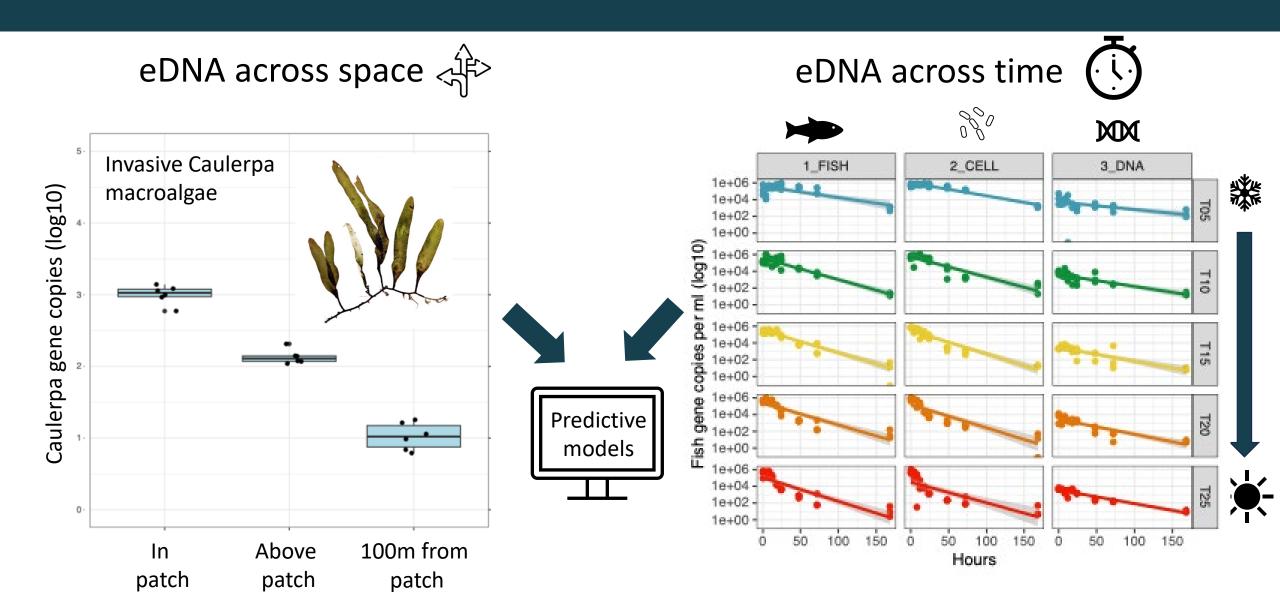
Research to advance eDNA method adoption



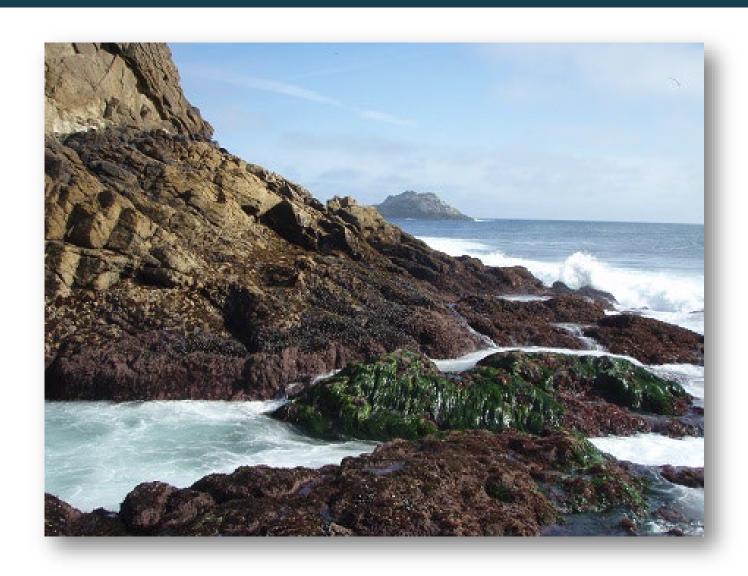




eDNA fate and decay



Expanding DNA Reference Libraries





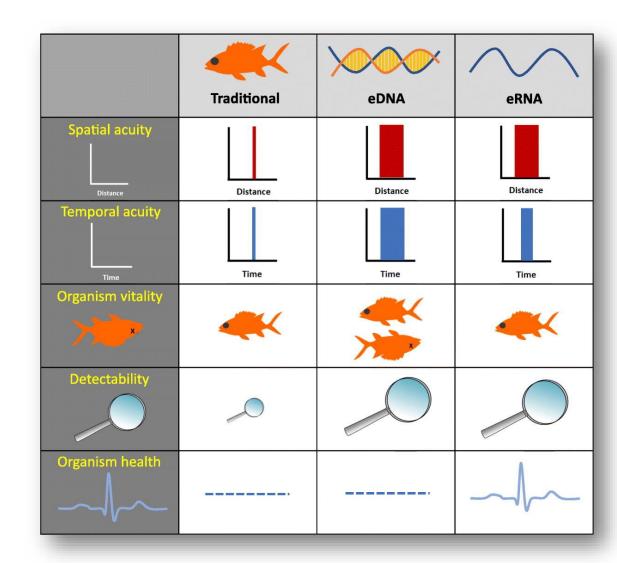




- California has allocated \$9M for Intertidal Biodiversity DNA Barcode Library
- Will help to advance our understanding of biodiversity hotspots and species range shifts
- SCCWRP is helping to lead the development of the standardized field and lab protocols

Live vs. dead (eRNA)

- eRNA degrades faster than eDNA and is only produced by living organisms
- Targeting eRNA will allow us to distinguish live vs. dead signals and also constrain timeframe of detections
- eRNA approaches have great potential for both our pathogen (e.g. Vibrio) as well as our harmful algal bloom monitoring



eDNA method standardization

California Molecular Methods Workgroup

Marine Technology Society eDNA Technology Committee

International eDNA Standardization Task Force











eDNA: research to implementation

- Environmental DNA research has advanced rapidly in the past 10 years
- Targeted species applications are leading the way for eDNA method adoption for routine implementation
- Community-based approaches are developing rapidly thanks to partner pilot studies and tool refinement
- eDNA method standardization is a key priority across all applications to ensure eDNA observations are robust, repeatability, and rock solid to support management decisions

